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(54) Title: BIOPOLYMER THICKENER

(57) Abstract: A novel strain of *Lactococcus lactis* subspecies *cremoris* ("Ropy 352") has been identified and isolated. Ropy 352 produces a previously unknown exopolysaccharide (EPS 352) that when expressed in or added to milk, imparts highly desirable sensory characteristics to the milk, including making the milk very thick, with a very smooth mouth-feel, and slightly sweet with an obvious "chewable-bite".



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BIOPOLYMER THICKENER

ACKNOWLEDGMENT OF GOVERNMENT SUPPORT

This invention was made in part with government support under The National Dairy Promotion and Research Board (i.e. Dairy Management Inc., DMI) and USDA/CSREES Special Research Grant. Accordingly the government has certain rights in this invention.

FIELD OF INVENTION

The field of the invention relates to biopolymers, enzymes that are contained within biopolymer synthesis pathways, nucleic acid sequences encoding such enzymes, and to organisms that make such biopolymers, wherein such biopolymers may be used to thicken liquids including liquid foods, as well as an additive to pharmaceuticals, beauty products, and coating agents.

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BACKGROUND

Microbial polysaccharides are used for a broad variety of industrial applications including food production, chemical production (e.g., detergents, cosmetics, paints, pesticides, fertilizers, flocculants, film formers, lubricants and explosives), pharmaceutical production and waste treatment. In food production, microbial polysaccharides are commonly used as thickening, gelling and homogenizing agents. When added to a liquid, microbial biopolymers contribute to viscosity, emulsion stabilization, surface tension and adhesiveness. Thickening applications are particularly important in the production of solid and semi-solid food products including dairy and non-dairy foods such as yogurt, buttermilk, salad dressings, cheese, and ice-cream. Thickening of liquid foods is desirable because of consumer preference for such thickened foods, which have a characteristic texture and "mouth feel." Thickening of liquid drinks is also desirable for use with elderly people who frequently have problems swallowing low-viscosity liquids (e.g., milk and fruit juices) due to an impaired swallowing reflex. The addition of thickener to such drinks facilitates swallowing and reduces aspiration of liquid into the trachea.

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Currently the only microbial polysaccharides used to any appreciable extent in industry are dextran, produced by *Leuconostoc mesenteroides*, xanthan gum, produced by *Xanthomonas campestris*, and gellan gum, produced by *Aureomonas elodea* ATCC31461 (Crescenzi, *Biotech. Prog.* 11:251-259, 1995). Xanthan gum was approved by the U.S. Food and Drug Administration (FDA) for use in foods in 1969. Today it is used in many foods such as bakery fillings, canned foods, frozen foods, pourable dressings, sauces, gravies, processed cheeses, and juice drinks. Xanthan gum is also used in oil recovery, pharmaceuticals, beauty products, and coating agents.

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Unfortunately, Xanthomonas campestris is a less than ideal source of polysaccharides for use in food production, since it is known to be pathogenic, and the biopolymer it produces has long been suspected of being pyrogenic (fever-inducing). Although xanthan gum is classified as "Generally Regarded as Safe" (GRAS) by the Food and Drug Administration (FDA), Xanthomonas campestris is not.

Lactic acid bacteria (LAB) are classified GRAS, and have been used for centuries in fermented dairy products such as yogurt, cheese, and sour-cream. A characteristic of some LAB in food production processes is their production of exopolysaccharides (EPS). EPS provide improved viscosity and mouth-feel while also preventing syneresis (separation) in fermented food products. Despite their ability to produce EPS, LAB are not generally used as sources of thickening agents (either within a milk-based culture or as a source of exogenous EPS) because the EPS-positive phenotype is readily lost (Dierkesen et al., *J. Dairy Sci.* 80(8):1528-1536, 1997). The LAB strain described in this disclosure stably produces EPS when cultivated on appropriate media.

SUMMARY OF THE DISCLOSURE

A natural isolate of *Lactococcus lactis*, named "*Lactococcus lactis* subspecies *cremoris* Ropy 352," hereinafter referred to simply as "Ropy 352", has been isolated. This strain contains a plasmid (EPS plasmid) that encodes at least 13 active genes (Figure 3). The enzymes encoded by these genes allow the bacteria to produce a previously unknown exopolysaccharide ("EPS 352"). Hence, in addition

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to providing EPS 352, the present invention also provides the nucleic acid sequences and the corresponding amino acid sequences of 13 of the open reading frames (ORFs; SEQ ID NO: 10) found on the EPS 352 plasmid.

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EPS 352, when expressed in or added to milk or other liquids, imparts desirable sensory characteristics to the milk, including making the milk very thick, with a very smooth mouth-feel, and slightly sweet with an obvious "chewable-bite." Ropy 352 producing EPS, or EPS 352 alone may be added to any milk-based or non milk-based product, including any liquid food product, to produce these sensory characteristics. In the Ropy 352 strain, the biosynthesis of EPS 352 is controlled by genes carried outside the chromosome on a plasmid of about 32 kb ("EPS 352 plasmid"). Precedent predicts that the EPS 352 genes are linked in an operon like fashion. The EPS 352 plasmid has been isolated from the Ropy 352 organism, and the plasmid has been transformed into a plasmid free nonropy laboratory strain of *Lactococcus*, MG1363. (Gasson, *J. Bacteriol.* 154:1-9, 1983.) The plasmid encoded EPS 352 genes are expressed in the transformed strain, producing a ropy EPS, which imparts desirable sensory characteristics (as detailed below) to milk-based media.

One aspect of the invention provides the isolated *Lactococcus lactis* subspecies *cremoris* Ropy 352 organism (Ropy 352) as deposited under the rules of the Budapest Treaty, USDA-ARS-NCAUR-NRRL deposit number NRRL B-30229. Ropy 352 can be added to liquids (e.g., solids, semi-solids and gels) to cause thickening. Such thickening is desirable for use in creating products such as food products, beauty care products, and pharmaceuticals. Additionally, the Ropy 352 organism can be used to produce food products by fermentation of a food substrate with a culture of the Ropy 352 organism. Accordingly, the invention also provides the products made through the addition of the Ropy 352 culture.

Another aspect of the invention provides the purified exopolysaccharide EPS 352. EPS 352 can be added to liquids to produce food products as well as other products such as pharmaceuticals. Examples of such liquids include, liquid food substrates, such as milk-based liquids, soy-based liquids, fruit juice, and whey-based liquids. Accordingly the invention also provides the products made through the addition of EPS 352.

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Yet another aspect of the invention provides the plasmid (contained in the deposited bacterial strain NRRL B-30229) that contains the open reading frames that encode the enzymes necessary for the production of EPS 352. This plasmid is approximately 32 kb in size. The identification of the plasmid allows for the production of EPS 352 by transgenic organisms that have been transformed with the EPS 352 plasmid. Furthermore, these transgenic organisms can be added to liquids to generate food products.

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Another aspect of the invention provides methods of using the individual enzymes encoded by the EPS 352 plasmid for the production of modified exopolysaccharides. Used in these methods the enzymes derived from the nucleic acid sequence of the EPS 352 plasmid can be combined with other genes that code for exopolysaccharide biosynthetic pathways enzymes such that the exopolysaccharide produced is distinct from that of the disclosed EPS 352. Furthermore, these methods can be practiced *in vitro* or *in vivo*. (Stingele et al., *Mol. Microbiol.* 32(6):1287-1295, 1999; Kranenburg et al., *J. Bacteriol.* 181(11):6347-6453, 1999; Stingele et al., *J. Bacteriol.* 181(20):6354-6360, 1999; and Klerrebezem et al., *Antonie van Leewenhoek* 76:357-365, 1999).

Another aspect of the invention provides methods of using EPS 352 in various pharmaceutical formulations. Used in this context EPS 352 can be incorporated dry into pill formulations or into liquids to increase the viscosity of the formulation and facilitate delivery of the active ingredients.

Another aspect of the invention provides methods of using EPS 352 in various beauty products, such as hair shampoos, hair bleaching compositions, hair conditioners, hair gels and mousse, skin creams, nail varnishes, facial foundation, skin tanning gels, hair removers, shaving creams and in pill coatings, children's products (i.e., crayons, non-toxic glues), in addition to various industrial processes. (Hilger et al., *J. Environ. Eng.* 125(12):1113, 1999 and Shah et al., *Appl. Biochem. Biotech.* 82(2):81, 1999.)

SEQUENCE LISTING

The nucleic and amino acid sequences listed in the accompanying sequence listing are shown using standard letter abbreviations for nucleotide bases, and three-

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letter code for amino acids. Only one strand of each nucleic acid sequence is shown, but the complementary strand is understood to be included by any reference to the displayed strand.

SEQ ID NO: 1 shows the nucleic acid sequence of a portion of the EPS 352 plasmid.

SEQ ID NO: 2 shows the amino acid sequence of the enzyme designated "R" in Figure 4, which is encoded by the nucleic acid sequence shown in SEQ ID NO: 1.

SEQ ID NO: 3 shows the amino acid sequence of the enzyme designated "X" in Figure 4, which is encoded by the nucleic acid sequence shown in SEQ ID NO: 1.

SEQ ID NO: 4 shows the amino acid sequence of the enzyme designated "A" in Figure 4, which is encoded by the nucleic acid sequence shown in SEQ ID NO: 1.

SEQ ID NO: 5 shows the amino acid sequence of the enzyme designated "B" in Figure 4, which is encoded by the nucleic acid sequence shown in SEQ ID NO: 1.

SEQ ID NO: 6 shows the amino acid sequence of the enzyme designated "C"

in Figure 4, which is encoded by the nucleic acid sequence shown in SEQ ID NO: 1.

SEQ ID NO: 7 shows the amino acid sequence of the enzyme designated "D" in Figure 4, which is encoded by the nucleic acid sequence shown in SEQ ID NO: 1.

SEQ ID NO: 8 shows the amino acid sequence of the enzyme designated "E"

in Figure 4, which is encoded by the nucleic acid sequence shown in SEQ ID NO: 1.

SEQ ID NO: 9 shows the amino acid sequence of the enzyme designated "O" in Figure 4, which is encoded by the nucleic acid sequence shown in SEQ ID NO: 1.

SEQ ID NO: 10 shows the amino acid sequence of the enzyme designated "P" in Figure 4, which is encoded by the nucleic acid sequence shown in SEQ ID NO: 1.

SEQ ID NO: 11 shows the amino acid sequence of the enzyme designated "F" in Figure 4, which is encoded by the nucleic acid sequence shown in SEQ ID NO: 1.

SEQ ID NO: 12 shows the nucleic acid sequence encoding Eps "M" and Eps "N."

SEQ ID NO: 13 shows the amino acid sequence of the enzyme designated "N" in Figure 4, which is encoded by the nucleic acid sequence shown in SEQ ID NO: 12.

SEQ ID NO: 14 shows the amino acid sequence of the enzyme designated "M" in Figure 4, which is encoded by the nucleic acid sequence shown in SEQ ID NO: 12.

SEQ ID NO: 15 shows the nucleic acid sequence encoding the enzyme designated "U."

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SEQ ID NO: 16 shows the amino acid sequence of Eps "U," which is encoded by SEQ ID NO: 15.

BRIEF DESCRIPTION OF THE DRAWINGS

10 Figure 1 describes the degree of phosphate protonation. As sodium hydroxide is added to the polysaccharide solution, there is only one inflection in the titration profiles, indicating that the phosphate group in the EPS 352 is in the form of a phosphodiester linkage rather than as the monoester, which would have shown 2 inflection points.

Figure 2 shows double stranded sequence data from the EPS 352 plasmid and the corresponding amino acid sequences named EpsM and EpsN. The insertion site of the ISS1 element is indicated in EspN and which confers a non-ropy phenotype in Ropy 352, thus linking these two open reading frames to EPS 352 expression.

Figure 3 shows the alignments of the ORF designated "N" in Figure 4 and the ORF designated "M" in Figure 4 to each other as well as to an enzyme (EpsG) involved in eps biosynthesis in *Lactococcus lactis* NIZOB40. The overall identity between ORF "M" and EpsG is 24% and between ORF "N" and EpsG is 25%.

Figure 4 is a diagram of the organization of the genes on the EPS 352 plasmid. The large arrows with letters inside represent genes and their orientation. The square with the letter X is a non-functional gene as it is missing its beginning (5' prime sequence). Eps ORFs are designated M, N, O, and P. The site of the ISS1 insertion, which disrupted EPS 352 production, is indicated by an downward pointing arrow that points to a position in Eps N.

Figure 5 shows the DNA and amino acid sequence of the entire EPS operon from upstream of the promoter to downstream of the terminator. This sequence is

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6850 bp in length. The starts of the open reading frames are labeled with the gene name (corresponding to Figure 4) printed in the right margin.

Figure 6 shows the nucleic acid sequence of Eps U. The start and stop codons are underlined.

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DETAILED DESCRIPTION

DEFINITIONS and ABBREVIATIONS

Unless otherwise noted, technical terms are used according to conventional usage. Definitions of common terms in molecular biology may be found in Benjamin Lewin, Genes VII, Oxford University Press, 1999 (ISBN 0-19-879276-X); 10 Kendrew et al. (eds.), The Encyclopedia of Molecular Biology Blackwell Science Ltd., 1994 (ISBN 0-632-02182-9); and Robert A. Meyers (ed.), *Molecular Biology* and Biotechnology: a Comprehensive Desk Reference, VCH Publishers, Inc., 1995 (ISBN 1-56081-569-8).

W/V means weight per unit volume.

15 kDa means kilodaltons.

MWCO means molecular weight cutoff

TCA means trichloroacetic acid.

Mol % means molar percent

mPA-s means millipascals

20 **n.d.** means none detected.

> Lactococcus lactis subspecies cremoris Ropy 352 ("Ropy 352") is the organism deposited under the Budapest Treaty as USDA-ARS-NCAUR-NRRL deposit number NRRL B-30229. Ropy 352 has the characteristic property of producing the exopolysaccharide EPS 352 under suitable growth conditions, e.g., streaked onto whey agar or defined lactococcal medium containing glucose agar plates and incubated at 30°C.

EPS 352 is an exopolysaccharide that is produced by Ropy 352 and that has the following characteristics:

Composition: Glucose:

range of 54% to 58%

Galactose:

range of 42% to 46%

Charged:

Yes

Molecular weight:

range of 800,000 to 8,000,000

-8-

(average of 1,600,000)

Phosphorous: Present in backbone or sidechain

Structure: Endpoints: galactose; Branchpoints: glucose

Several gene products are required for EPS 352 biosynthesis. The EPS biosynthetic genes are located extrachromasomally on the EPS 352 plasmid. Precedent indicates that these genes are organized in an operon like fashion.

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EPS 352 plasmid is an extrachromosomal plasmid of approximately 32 kb in size that carries the EPS 352 biosynthetic genes. Current methods used to estimate plasmid size are not exact. For instance, the perceived size of a plasmid may be effected by the degree of relaxation of the plasmid and the degree to which proteins may be associated with the plasmid. Thus, the EPS 352 plasmid is believed to be about 32 kb in size, and may be, for example, from 30 to 38 kb in size. Several research groups have linked EPS biosynthesis with plasmids of various sizes: 6.8 kb, 25.8 kb, 28 kb, 40.2 kb, and 45.5 kb (Vescovo et al., *Biotech. Letters II* 10:709-712, 1989; Neve et al., *Biochimie* 70:437-442, 1988; Vedamuthu et al., *Appl. Environ. Microbiol.* 51:677-682, 1986; Kranenburg et al. *Mol. Microbiol.* 24:387-397, 1997; and Von Wright et al., *Appl. Environ. Microbiol.* 53:1385-1386, 1987).

Food means any eatable or drinkable substance consumed by humans or animals, e.g., milk, cream, dairy products, soy products, fruit juice, vegetable juices, ice cream, soups, etc.

Food Product means any food that is produced by altering its original state, e.g., milk to which has been added EPS 352.

Milk is used broadly herein to include all dairy products regardless of fat content or lactose content. The term as used herein also includes substances commonly used in place of milk, such as soy used as "soy milk". The term also includes milk products from animals other than cows, including goat milk.

Liquid as used herein includes fluids with varying degrees of fluidity including highly fluid liquids such as non-fat milk, thicker liquids such as full fat milk and cream, semi-solid substances, and gels such as yogurt and other fermented milk products. A liquid may be altered from its original state to produce an altered liquid, e.g., an adhesive solution, a paint emulsion, a lubricant, or a fruit juice to which EPS 352 has been added.

A Milk-Based liquid is any liquid wherein milk forms an appreciable percentage of the total volume of the liquid. For example, a liquid having 0.10% or more of milk solids.

A Soy-Based liquid is any liquid wherein soy forms an appreciable percentage of the total volume of the liquid. For example, a liquid having 0.10% or more of soy solids

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To Thicken means to decrease fluidity and increase viscosity.

Thickener means any substance used to thicken, including, for instance, exopolysaccharides. A thickener may be produced by organisms cultured within a medium or may be added exogenously to a medium.

Mouth-feel is a term of art used in the food industry to describe sensory characteristics of a food. It has the same meaning as the word "texture" which has been previously defined as "the composite of the structural elements of the food and the manner in which it registers with the physiological sense" (Szczesniak, *J. Food Science* 28:385-389, 1963), or "the composite of those properties which arise from the physical structural elements and the manner in which it registers with the physiological senses" (Sherman, *J. Food Science* 27:381-385, 1970).

Pharmaceutical a chemical compound or composition capable of inducing a desired therapeutic or prophylactic effect when properly administered to a subject.

Beauty care product is an externally applied product that is intended to alter the appearance of the subject to which it has been applied.

Coating agent an agent applied to the exterior surface of an object. A coating agent generally forms a thin layer on the surface of the object.

Transformed refers to a cell into which a nucleic acid molecule has been introduced by molecular biology techniques. The term encompasses all techniques by which a nucleic acid molecule might be introduced into such a cell, including transformation with plasmid vectors, transfection with viral vectors, and introduction of naked DNA by electroporation, lipofection, and particle gun acceleration.

Purified does not require absolute purity; rather, it is intended as a relative term. Thus, for example, a purified polysaccharide preparation is one in which the subject polysaccharide is more pure than in its natural environment within a cell or within a cell culture medium. Generally, a polysaccharide preparation is purified

such that the polysaccharide represents at least 50% of the total polysaccharide content of the preparation.

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Isolated an *isolated* nucleic acid has been substantially separated or purified away from other nucleic acid sequences in the cell of the organism in which the nucleic acid naturally occurs, i.e., other chromosomal and extrachromosomal DNA and RNA. The term "isolated" thus encompasses nucleic acids purified by standard nucleic acid purification methods. The term also embraces nucleic acids prepared by recombinant expression in a host cell, as well as chemically synthesized nucleic acids.

ORF is an open reading frame. An ORF is a contiguous series of nucleotide triplets coding for amino acids. These sequences are usually translatable into a peptide.

Operably linked means a first nucleic acid sequence is operably linked with a second nucleic acid sequence when the first nucleic acid sequence is placed in a functional relationship with the second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Generally, operably linked DNA sequences are contiguous and, where necessary to join two protein coding regions, in the same reading frame.

Probe is an isolated nucleic acid attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes.

Target Nucleic Acid is a nucleic acid that hybridizes with a probe. The conditions under which hybridization occurs may vary with the size and sequence of the probe and the target sequence.

By way of illustration, only a hybridization experiment may be performed by hybridization of a DNA probe (for example, a probe derived from the EPS 352 plasmid labeled with a chemiluminescent agent) to a target DNA molecule which has been electrophoresed in an agarose gel and transferred to a nitrocellulose membrane by Southern blotting (a technique well known in the art and described in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd ed., vols. 1-3, Cold Spring Harbor, New York, 1989).

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Hybridization with a radio-labeled probe is generally carried out in a solution of high ionic strength such as 6 x SSC at a temperature that is 20°C-25°C below the melting temperature, T_m, described below. For such Southern hybridization experiments where the target DNA molecule on the Southern blot contains 10 ng of DNA or more, hybridization is typically carried out for 6-8 hours using 1-2 ng/mL radiolabeled probe. Following hybridization, the nitrocellulose filter is washed to remove background hybridization. The wash conditions should be as stringent as possible to remove background hybridization but to retain a specific hybridization signal. The term T_m represents the temperature above which, under the prevailing ionic conditions, the radiolabeled probe molecule will not hybridize to its target DNA molecule. The T_m of such a hybrid molecule may be estimated from the following equation:

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 $T_m = 81.5$ °C - 16.6 ($\log_{10} [Na^+]$)+0.41 (%G+C) - 0.63 (% formamide) -(600 / 1) Where I = the length of the hybrid in base pairs. This equation is valid for concentrations of Na^+ in the range of 0.01M to 0.4M, and it is less accurate for calculations of T_m in solutions of higher [Na^+]. The equation is primarily valid for DNAs whose G+C content is in the range of 30% to 75%, and applies to hybrids greater than 100 nucleotides in length (the behavior of oligonucleotide probes is described in detail in Ch. 11 of Sambrook et al., 1989).

Generally hybridization wash conditions are classified into categories, for example very high stringency, high stringency, and low stringency. The conditions corresponding to these categories are provided below.

Very High Stringency (detects sequences that share 90% sequence identity) 25 Hybridization in 65°C 16 hours 5xSSC at Wash twice 2xSSC in at Room temp. 15 minutes each Wash twice in 0.2xSSC 65°C at 20 minutes each

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High Stringency (detects sequences that share 80% sequence identity	or
greater)	

Hybridization	in	3x	SSC	at	65°C	16 hours
Wash twice	in	2x	SSC	at	Room temp.	15 minutes each
Wash twice	in	0.5x	SSC	at	55°C	20 minutes each

Low Stringency (detects sequences that share greater than 50% sequence identity)

	Hybridization	in	3x	SSC	at	65°C	16 hours
10	Wash twice	in	2x	SSC	at	Room temp.	20 minutes

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The above example is given entirely by way of theoretical illustration. One skilled in the art will appreciate that other hybridization techniques may be utilized and that variations in experimental conditions will necessitate alternative calculations for stringency.

Conservative amino acid substitutions are those substitutions that, when made, least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids that may be substituted for an original amino acid in a protein and that are regarded as conservative substitutions.

TABLE 1

Original Residue	Conservative		
	Substitutions		
ala	ser		
arg	lys		
asn	gln; his		
asp	glu		
cys	ser		
gln	asn		
glu	asp		
gly	pro		

Conservative		
Substitutions		
asn; gln		
leu; val		
ile; val		
arg; gln; glu		
leu; ile		
met; leu; tyr		
thr		
ser		
tyr		
trp; phe		
ile; leu		

Conservative substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side chain.

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The substitutions which in general are expected to produce the greatest changes in protein properties will be non-conservative. For instance, changes in which (a) a hydrophilic residue, e.g., seryl or threonyl, is substituted for (or by) a hydrophobic residue, e.g., leucyl, isoleucyl, phenylalanyl, valyl or alanyl; (b) a cysteine or proline is substituted for (or by) any other residue; (c) a residue having an electropositive side chain, e.g., lysyl, arginyl, or histadyl, is substituted for (or by) an electronegative residue, e.g., glutamyl or aspartyl; or (d) a residue having a bulky side chain, e.g., phenylalanine, is substituted for (or by) one not having a side chain, e.g., glycine.

Primers are short nucleic acids, preferably DNA oligonucleotides 10 nucleotides or more in length, which are annealed to a complementary target DNA strand by nucleic acid hybridization to form a hybrid between the primer and the target DNA strand, then extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR) or other nucleic-acid amplification methods known in the art.

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 30, 40, 50, 60, 70, 80, 90, 100, or 150 consecutive nucleotides of the disclosed nucleic acid sequences.

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Methods for preparing and using probes and primers are described in the references, for example Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd ed., vol. 1-3, Cold Spring Harbor, New York, 1989; Ausubel et al., *Current Protocols in Molecular Biology*, Greene Publ. Assoc. & Wiley-Intersciences, 1987; Innis et al., *PCR Protocols, A Guide to Methods and Applications*, 1990. PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as *Primer* (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge, MA).

Recombinant nucleic acid is a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook et al. (1989). The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector, used to transform a cell.

Sequence identity: The similarity between two nucleic acid sequences or between two amino acid sequences is expressed in terms of the level of sequence identity shared between the sequences. Sequence identity is typically expressed in terms of percentage identity; the higher the percentage, the more similar the two sequences.

Methods for aligning sequences for comparison are well known in the art. Various programs and alignment algorithms are described in: Smith & Waterman, Adv. Appl. Math. 2:482, 1981; Needleman & Wunsch, J. Mol. Biol. 48:443, 1970; Pearson & Lipman, Proc. Natl. Acad. Sci. USA 85:2444, 1988; Higgins & Sharp,

Gene 73:237-244, 1988; Higgins & Sharp, CABIOS 5:151-153, 1989; Corpet et al., Nucleic Acids Research 16:10881-10890, 1988; Huang, et al., CABIOS 8:155-165, 1992; and Pearson et al., Methods in Molecular Biology 24:307-331, 1994. Altschul et al., J. Mol. Biol. 215:403-410, 1990, presents a detailed consideration of sequence alignment methods and homology calculations.

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The NCBI Basic Local Alignment Search Tool (BLASTTM) (Altschul et al., J. Mol. Biol. 215:403-410, 1990 is available from several sources, including the National Center for Biotechnology Information (NBCI, Bethesda, MD) and on the Internet, for use in connection with the sequence analysis programs blastn, blastn, blastx, tblastn and tblastx. BLASTTM can be accessed on the interned at NBCI website. A description of how to determine sequence identity using this program is available at the web site. As used herein, sequence identity is commonly determined with the BLASTTM software set to default parameters. For instance, blastn (version 2.0) software may be used to determine sequence identity between two nucleic acid sequences using default parameters (expect = 10, matrix = BLOSUM62, filter = DUST (Tatusov and Lipmann, in preparation as of December 1, 1999; and Hancock and Armstrong, Comput. Appl. Biosci. 10:67-70, 1994), gap existence cost = 11, per residue gap cost = 1, and lambda ratio = 0.85). For comparison of two polypeptides, blastp (version 2.0) software may be used with default parameters (expect 10, filter = SEG (Wootton and Federhen, Computers in Chemistry 17:149-163, 1993), matrix = BLOSUM62, gap existence cost = 11, per residue gap cost = 1, lambda = 0.85).

For comparisons of amino acid sequences of greater than about 30 amino acids, the "Blast 2 sequences" function of the BLASTTM program is employed using the default BLOSUM62 matrix set to default parameters, (gap existence cost of 11, and a per residue gap cost of 1). When aligning short peptides (fewer than around 30 amino acids), the alignment should be performed using the Blast 2 sequences function, employing the PAM30 matrix set to default parameters (open gap 9, extension gap 1 penalties). Proteins with even greater similarity to the reference sequences will show increasing percentage identities when assessed by this method, such as at least 45%, at least 50%, at least 60%, at least 80%, at least 85%, at least 90%, or at least 95% sequence identity.

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METHODS

General Methods

The present invention utilizes standard laboratory practices for the cloning, manipulation and sequencing of nucleic acids, purification and analysis of proteins and other molecular biological and biochemical techniques, unless otherwise stipulated. Such techniques are explained in detail in standard laboratory manuals such as Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd ed., vol. 1-3, Cold Spring Harbor, New York, 1989; and Ausubel et al., Current Protocols in Molecular Biology, Greene Publ. Assoc. & Wiley-Intersciences, 1989. Other techniques specific to Lactococcus are discussed in the inventors' publications including: Dierksen et al., *Genetics of Streptococci, Enterococci and Lactococi*, (Ferretti et al., eds.), 1995; Basel, *Dev. Biol. Stand* 85:469-480, 1995; Dierksen et al., *J. Dairy Sci.*, 80(8):1528-1536, 1997; and Knoshaug et al., *J. Dairy Sci.* 83:633-640, 2000.

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1. Growth and Characterization of the Ropy 352 organism.

The EPS 352 producing organism, Lactococcus lactis subspecies cremoris Ropy 352, was isolated, classified and deposited under the Budapest Convention as USDA-ARS-NCAUR-NRRL deposit number NRRL B-30229. Ropy 352 may be obtained on demand from the USDA-ARS-NCAUR-NRRL at Agricultural Research Service Culture Collection (NRRL), National Center for Agricultural Utilization Research (NCAUR), Agricultural Research Service (ARS), U.S. Department of Agriculture (USDA), 1815 North University Street, Peoria, IL 61604 U.S.A. Ropy 352 was streaked onto whey agar or defined lactococcal media containing glucose (DLMG) agar. Whey agar (Vedamuthu et al., Appl. Microbiol. 51:677-682, 1986) made as previously described with the following modifications: yeast extract (5 g, Difco Laboratories, Detroit, MI) and sodium β-glycerophosphate (19 g, Sigma Chemical Co., St. Louis, MO) were added to the centrifuged supernatant and the volume brought up to 600 mL. The second part of the media consisted of 15 g of agar and 3 drops of antifoam A (Sigma) in 400 mL of water. Both portions were autoclaved for 12 min, removed promptly, cooled to 50°C, mixed, and poured into sterile petri plates. DLMG agar (Molenaar et al., J. Bacteriol. 175:5438-5444.

1993.) was prepared as two parts; part one consisted of the base media which was prepared in 758 mL of water, heated to dissolve the components, mixed with 10 mL of the metals, vitamins, and nucleic acid solutions and 12 mL of 20% glucose or lactose solution, filter sterilized, and heated to 55°C in a water bath. Part two consisted of 10 g of agar and 2 drops of antifoam A (Sigma) which were mixed into 200 mL of water, autoclaved, and cooled to 55°C. Part one was mixed into part two and poured into sterile petri plates. Ropy 352 was streaked onto plates and incubated at 30°C to produce macroscopic, individual, EPS 352 producing colonies of Ropy 352 (procedure described in inventors' publications listed above).

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The EPS 352 may be recognized by the formation of viscous ropes greater than five mm in length originating from a whey agar or DLMG agar. Whey agar plates were incubated at 30°C for 48 h. Characteristic ropy phenotype is apparent from viscous rope greater than 5 mm formed when a colony is touched with a sterile toothpick. These ropes became visible when the colony was touched with a sterile toothpick and the toothpick was drawn away from the colony, thus, stretching the EPS 352 out. An additional way to recognize EPS 352 is by the formation of viscous ropes in liquid milk inoculated with Ropy 352 organism. Liquid milk was sterilized by steaming for 30 min and 10 mL of milk were inoculated with 0.5 mL of an overnight Ropy 352 culture. The milk was incubated for 18 hours at 30°C and visually examined for ropy EPS expression. These viscous ropes were visualized by touching the milk with a toothpick and drawing the toothpick away from the milk.

2. Purification and Characterization of EPS 352.

An individual EPS 352 producing Ropy 352 colony from a whey agar plate was picked and used to inoculate 1 L of polysaccharide production medium in a 2.8 L Fernbach flask. The medium was cultured at 30°C for 16 to 20 hours without shaking. The polysaccharide production medium consisted of 10% w/v nonfat milk in water, which was prepared by stirring 100 g dry milk powder into 1 L deionized water at room temperature for 1 hour and then sterilizing the mixture in an autoclave for 12 minutes at 120°C.

Ropy 352 culture broths were transferred to 500 mL centrifuge bottles and insoluble fractions were pelleted at 10 K x g for 20 minutes. Clarified supernatants

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were dialyzed (6-8 kDa MWCO, Spectra/Por 1; Spectrum Laboratories, Inc., Laguna Hills, CA) against water containing 0.02% sodium azide for at least 24 hours.

An equal volume of absolute ethanol was added to the contents of the dialysis tubing and stirred in an ice bath. Ropy 352 cultures formed a precipitate of elongated ropes that were collected by centrifugation as described above. This was termed the Ropy fraction and contained EPS 352.

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From 1 L of 10% nonfat milk medium, 34 mg of total polysaccharide was recovered from Ropy 352 cultures after centrifugation and dialysis. The polysaccharide responsible for the ropy characteristic (EPS 352) was purified by precipitation with 50% ethanol, followed by trichloroacetic acid (TCA) removal of residual protein. This Ropy fraction contained 10 mg of polysaccharide and was essentially protein free (<20 µg/mg in the final product). The Ropy fraction also contained 2.3 µg phosphorus/mg polysaccharide.

Compositional analysis of EPS 352 revealed a repeating structure composed of approximately 54% to 58% glucose, and 42% to 46% galactose. Compositional data suggests a novel structure for EPS 352 with glucose as the branch residue and galactose located at the end points.

The predominant sugar found in EPS 352, at 36 mol%, is (1,4)-linked glucose. The only sugar found as terminal non-reducing end groups (i.e., had a single linkage position) was galactose at 27 mol%; this quantity is indicative of a highly branched structure. A (1,4,6)-linked glucose reside was found at a concentration of 21 mol%; the three linkage sites indicate that it is a branch point in this structure. The least represented sugar was the (1,4)-linked galactose, which occurred at a concentration of 15 mol%. Results from this analysis are listed in Table 2:

Table 2
Identification of permethylated PAAN (Peracetylated aldononitrile)
derivatives from Ropy 352 and Ropy polysaccharides

		Ropy fraction from Ropy
PAAN methyl sugar	Linkage site	352 (mol%)
2,3,4,6-tetra-O-methyl galactose	1	27
2,3,6-tri-O-methyl galactose	1,4	15
2,4,6-tri-O-methyl galactose	1,6	n.d.
		(none detected)
2,3,4-tri-O-methyl galactose	1,6	n.d.
2,3,6-tri-O-methyl glucose	1,4	36
2,3,4-tri-O-methyl glucose	1,6	n.d.
3,4,6-tri-O-methyl mannose	1,2	n.d.
2,3-di-O-methyl glucose	1,4,6	21
3,4-di-O-methyl glucose	1,2,6	n.d.
2,4-di-O-methyl mannose	1,3,6	n.d.

The degree of phosphate protonation is shown in Figure 1. As sodium hydroxide was added to the polysaccharide solution, there was only one inflection in the titration profiles, indicating that the phosphate group in the Ropy fraction polysaccharides is in the form of a phosphodiester linkage rather than as the monoester, which would have shown 2 inflection points.

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3. Viscosity of Milk Culture During 25 hour Fermentation with Ropy 352.

1 L of milk was inoculated with a single whey agar-grown colony of Ropy 352. Viscosity was measured with a Brookfield model LVTDV-I digital viscometer (Stoughton, MA) using a LV1 spindle.

The viscosity of the Ropy 352 culture reached a value of 44000 mPA-s at 24 hours, compared to an initial viscosity of 1 mPa-s (see Table 3). This data verifies the phenotypic observation that Ropy 352 culture thickens a liquid food product (milk).

Table 3
Viscosity change (in mPa-s) after 24 h

Strain	Sample	0 h	24 h
Ropy 352	Fermented milk	1.0	44000
No cells	Milk	1.0	1.0

4. Isolation and Characterization of the Biosynthetic EPS 352 Plasmid.

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The EPS 352 plasmid is a plasmid of about 32 kb in size that may be isolated from Ropy 352. A 2.2 KB fragment from the EPS 352 plasmid (Figure 2) and a 6.85 kb fragment (Figure 4) have been sequenced. These sequences encodes ORFs M and N which show homology to a class of sugar transfer enzymes (glycosyltransferases) known to be involved in EPS biosynthesis (Figure 2). Several restriction endonucleases cut this plasmid, including *EcoRI*, *EcoRV*, *HindIII*, *SacI*, *SphI*, *DraI*, *HincII*, *NdeI*, *Sau3AI*, and *SpeI*.

The EPS 352 plasmid contains all biosynthetic genes coding for the enzymes needed to make EPS 352. This was demonstrated by the following experiment. The EPS 352 plasmid, containing an erythromycin resistant encoded insertion element for selection, was isolated from a culture of Ropy 352 using DNA preparation methods as described in Knoshaug et al., J. Dairy Science 83:633-640, 2000. (Ref for plasmid DNA isolation: O'Sullivan et al., Appl Environ Microbiol. 59:2730-2733, 1993). This DNA was used to transform a plasmid-free nonropy lactococcal strain, MG1363 by electroporation as described (Dornan et al., Lett. Appl. Microbiol. 11:62-64, 1990; Holo et al., Appl. Environ. Microbiol. 55:3119-3123, 1989). Cells were grown for 24 hours in M17-glucose media supplemented with 0.3 M sucrose and 2% (MG1363) or 0.5% (Ropy352) glycine. Cells were pelleted, washed in cold 0.3 M sucrose three times, and resuspended in 200 µl of 0.3 cold M sucrose. DNA was added to the cells and the mixture was transferred to a chilled electroporation cuvette (0.2 cm gap). The cells were shocked (2.5 kV, 200 ohms, 25 µF) and resuspended in 8 mL of growth media supplemented with 0.3 M sucrose and 50 ng/mL em. Cells were allowed to recover for 1.5 hours before plating on whey agar containing 2 µg/mL em. Erythromycin resistant transformants were selected, and then screened for the ropy EPS 352 phenotype. MG1363 containing the EPS 352 plasmid was analyzed by Southern blot to verify the presence of the plasmid. The probe used was 1.6 kb long and specific to the Ropy 352 EPS ORF M and ORF N genes. Results demonstrated that the probe reacted with a 32 kb plasmid in Ropy352 (un-nicked and nicked forms) and with a 37 kb plasmid in EK356 (EPS

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352 plasmid containing a 5.4 kb erythromycin resistant encoded insertion element for selection; un-nicked and nicked forms).

The southern blot analysis was additionally confirmed by testing the transformed bacteria for the Ropy phenotype. Results showed that the phenotypic carried over to the MG1363 strain.

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5. Production of Food Products by Adding EPS 352 to a Food Substrate.

EPS 352 can be added to a liquid food substrate to increase viscosity and thickness of the liquid and to enhance texture and mouth-feel. Liquid food substrates may include, but are not limited to: milk (including low-fat and non-fat milk), milk-based liquids, whey-based liquids, soy-based liquids, fruit-juices, and oil-based liquids and emulsions. EPS 352 can be used to enhance the thickness and texture of, for example, yogurt, milk-shakes, fruit-juices, soy drinks, Scandinavian fermented milk products (e.g., "villi, "langfil," and "filmjolk,"), bakery fillings, dressings, sauces and gravies. EPS 352 can also be added to solid or semi-solid food substrates to enhance the texture of, for instance, frozen foods, canned foods and cheeses. Thickness of the liquid food substrate will increase in proportion to the amount of EPS 352 added. EPS 352 may be added to any liquid food substrate in an amount necessary to produce the desired consistency. Determining an amount necessary to produce a desired consistency is a simple matter of empirical experimentation.

A specific example of a food product made using EPS 352 is a thickened, non-fermented food product that has the qualities of yogurt, but without the need for fermentation. Milk (e.g., non-fat milk) can be used as a liquid food substrate to which an amount of EPS 352 can be added, sufficient to cause thickening to a desired consistency. EPS 352 may be supplied in the form of an essentially pure powder and added directly to the milk. The powder may be mixed into the milk at room temperature using conventional methods and the mixture may then be aliquoted into sealed containers and pasteurized. Such a product would be low in fat, have a yogurt-like consistency, and would not require fermentation, a step which is time-consuming, expensive and prone to microbial contamination.

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6. Production of Milk-Derived Fermented Food Products by Adding a Pure Culture of the Ropy 352 Organism to a Food Substrate and Fermenting the Mixture.

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Ropy 352 can be used to produce fermented food products such as yogurt (and other products as listed above). Such products are described as probiotic (this refers to organisms who are ingested, such as the LAB, which contribute to the health and balance of the human's intestinal tract thus possibly protecting against disease and improving nutrition). During fermentation, Ropy 352 produces the EPS 352 exopolysaccharide which imparts desirable qualities to certain foods. In particular, EPS 352 gives fermented milk products a very smooth, rich mouth-feel with a slightly sweet flavor.

A specific example of a fermented food product made using Ropy 352 is yogurt. Milk (e.g., either whole, 2% or non-fat milk) can be used as a liquid food substrate to which a pure culture of Ropy 352 can be added. The culture may be fermented, for instance at 30°C without shaking for 16 to 20 hours. The EPS 352 culture may be supplied in the form as an aliquot of liquid culture or an inoculum from an agar plate (such as milk or whey agar plate). Following fermentation, the fermented product may be aliquoted into sealed containers and pasteurized. A second specific example of a fermented food product made using Ropy 352 is a power shake for the elderly and diet shakes for the obese. Trade names such as SlimfastTM or EnsureTM can be used as a liquid food substrate to which a pure culture of Ropy 352 can be added. Both SlimfastTM and EnsureTM were inoculated with a culture of Ropy352 and incubated at 30°C for 24 hours, respectively. The results showed that not only did Ropy 352 thicken these products, but it also added active culture (probiotic) status.

The duration and temperature of fermentation may vary. Representative temperatures may range from about 17°C to 30°C and duration of fermentation of a batch culture may be from about 10 to 36 hours. Alternatively, fermentation may be done as a continuous culture with portions of the fermented product being periodically removed.

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7. The Use of Enzymes Derived from the EPS 352 Plasmid

Enzymes derived from the EPS 352 plasmid can be used either *in vitro* or *in vivo* to produce and or modify EPS structure. Furthermore, these enzymes can be modified through the inclusion of one or more conservative amino acid substitutions, however, such conservative amino acid substituted variants will continue to maintain the same activity of the enzyme from which they are derived.

a. in vitro

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Enzymes from the EPS 352 plasmid can be combined with other enzymes and substrates in vivo, such that an EPS is produced with the desired characteristics. *In vitro* production of an EPS involves provide the isolated enzymes that are to be used in the synthesis as well as the various substrates necessary for the production of the EPS. Detailed examples of EPS production *in vitro* are well known in the art and can be found for example in Bossia et al., *Cell Mol Biol (Noisy-le-grand)*42(5):737-58, 1996 and

15 Semino et al., *J Gen Microbiol* **139** (Pt 11):2745-56, 1993.

b. in vivo

The enzymes produced from the expression of ORFs, such as ORF M (SEQ ID NO: 14), ORF N (SEQ ID NO: 13), ORF O (SEQ ID NO: 9), and ORF P (SEQ ID NO: 10) that are derived from the EPS 352 plasmid can be placed under the control of heterologous control sequences. Such control sequences can be selected from constituative promoters, inducible promoters, enhancers, and various terminators. Together the control sequence(s) operably linked to the ORF is termed the "transgene". The transgene can then be transformed into a host organism that supports the production of an EPS. Upon expression of the protein from the transgene at least a portion of the EPS generated from the transformed host organism will be distinct from the non-transformed host organism.

It is also possible that the control sequences found in the EPS 352 plasmid can be used to express one of more of the ORF from the EPS 352 plasmid. Used in this way the "transgene" generated will be the result of using recombinant DNA technology to manipulate the endogenous EPS 352 plasmid such that the naturally occurring EPS 352 plasmid is not intact. Such transgenes result from the introduction of additional copies of one or more of the ORFs that are in the naturally

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occurring EPS 352 plasmid. It is also possible that enzymes from other EPS producing organisms will be introduced into the EPS 352 operon such that the host cell expresses an EPS that is distinct from the Ropy 352 disclosed herein.

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EXAMPLES

1. Production of a Thickened Milk Product by Adding a Pure Culture of the Ropy 352 Organism to Milk and Fermenting the Mixture.

Ropy EPS 352 was expressed on plates containing whey agar and in liquid milk. The whey agar plates were incubated at 30°C for 48 hours. Colonies were then touched with a sterile toothpick to test for Ropy EPS 352 expression. Liquid milk was sterilized by steaming for 30 minutes. 10 mL of the sterilized milk were then inoculated with 0.5 mL of an overnight pure culture of the Ropy 352 organism. The milk was incubated for 18 hours at 30°C and visually examined for coagulation and ropy EPS 352 expression. Ropiness was indicated using a sterile glass rod to pull ropes from the milk.

2. Production of a Thickened Liquid Product by Adding a Pure Culture of the Ropy 352 Organism to Power Drinks Designed for the Elderly and Diet Drinks Designed for the Obese.

Ropy 352 was grown and EPS 352 was expressed in Slim FastTM (Slim-Fast Foods Co., West Palm Beach, Florida) chocolate diet drink and EnsureTM (Abbott Laboratories, Abbott Park, Illinois) chocolate fortified drink. Slim FastTM and EnsureTM drinks were inoculated with Ropy 352 and incubated for 18 hours at 30°C and visually examined for coagulation and ropy EPS 352 expression. Ropiness was determined using a sterile glass rod to pull ropes from the milk, and by visually examining how the fermented liquid poured from a flask.

3. Use of the EPS 352 Plasmid to Transform Cells and to Produce EPS 352.

The EPS 352 plasmid, containing an erythromycin resistant encoded insertion element for detection, was isolated from a culture of Ropy 352 using DNA preparation methods as described in Knoshaug et al., *J. Dairy Sci.* 83:633-640, 2000 (and as referred to in the methods section of this document). This DNA was used to

transform a plasmid-free nonropy lactococcal strain, MG1363. Erythromycin resistant transformants were selected, and then screened for the ropy EPS 352 phenotype. Those displaying the ropy EPS 352 phenotype were Gram stained to verify that Gram positive cocci were present. MG1363 containing the EPS 352 plasmid was analyzed by Southern blot to verify the presence of EPS 352 plasmid. Presence of the EPS 352 plasmid in MG1363 correlated to the acquisition of the ropy EPS 352 phenotype.

4. Use of EPS 352 as a Substitute for Xanthan Gum

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Xanthan gum is a high molecular weight polysaccharide derived from *Xanthomonas Campestris*. It contains D-glucose, D-mannose, and D-glucuronic acid as the dominant hexose units. For a more detailed discussion of the composition, physical and chemical properties, preparation, etc. of xanthan gum, see the following publications: Federal Register, Vol. 34, No. 53, Mar. 19, 1969, Subchapter B, Part 121, Subpart D; Keltrol, Technical Bulletin DB No. 18, Kelco Company, Clark, New Jersey.

Xanthan gum is currently used in a variety of compounds, as is evidenced by the fact that a search of the United States Patent and Trademark Office website on the Internet for "xanthan gum" in the claims of U.S. patents that have issued since 1976 identified 1,276 patents. These patents show xanthan gum being used in sprayable cleaning compositions (U.S. patent No. 5,948,743), hair conditioning shampoo (U.S. patent No. 5948,739), ballpoint pen ink (U.S. patent No. 5,925,175), time-specific controlled release dosage formulations (U.S. patent No. 5,891,474), to improve gloss retention of surfactants (U.S. patent No. 5,877,142), as wells as for many other purposes.

5. Enzymatic Activity of the Enzymes Produced By the EPS 352 Plasmid

The EPS plasmid contains at least 5 previously unidentified open reading frames encoding 5 previously unidentified enzymes (O, P, N, M, and U, which are provided in SEQ ID NOS: 9, 10, 12, 13, and 14, respectively). Sequence analysis using BlastTM searching indicates that the "M" enzyme (SEQ ID NO: 13) is a glycosyltransferase enzyme. Methods of testing glycosyltransferase activity are

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well known in the art and described in: van Kranenburg et al., *J. Bacteriol*. **181**(1):338-340, 1999; Kranenburg et al., *J. Bacteriol*. **181**(11):6347-6353, 1999; Stingele et al., *J.*

Bacteriol. 181(20):6354-6360, 1999; Kolkman et al., J. Bacteriol. 178(13):3736-3741 1996; Kolkman et al., J. Biol. Chem. 272(31):19502-19508; Breton, et al., Curr. Opin. Struct. Biol. 9:563-571, 1999; and Griffiths et al., J. Biol. Chem. 273(19):11752-11757, 1998, which are herein incorporated by reference.

Similarly, sequence analysis using BlastTM searching indicates that the "P" enzyme (SEQ ID NO: 10) is a polysaccharide polymerase. Methods of testing polysaccharide polymerase activity are well known in the art and described in: Gonzalez et al., *Proc.Natl. Acad. Sci.* 95:13477-13482, 1998; Stevenson et al., *J. Bacteriol.*

178(16):4885-4893, 1996; and Glucksmann et al., *J. Bacteriol.* **175**(21):7045-7055, 1993, which are herein incorporated by reference.

Sequence analysis using BlastTM searching indicates that the "N" enzyme (SEQ ID NO: 12) is a galactosyltransferase enzyme. Methods of testing galactosyltransferase activity are well known in the art and described in: van Kranenburg et al., *J. Bacteriol*.

181(1):338-340, 1999; Kranenburg et al., J. Bacteriol. 181(11):6347-6353, 1999;
Stingele et al., J. Bacteriol. 181(20):6354-6360, 1999; Kolkman et al., J. Bacteriol.
178(13):3736-3741, 1996; Kolkman, et al., J. Biol. Chem. 272(31):19502-19508,
1997; Breton et al., Curr. Opin. Struct. Biol. 9:563-571, 1999; and Griffiths et al., J. Biol. Chem. 273(19):11752-11757, 1998, which are herein incorporated by reference.

Sequence analysis using Blast[™] searching indicates that the "O" enzyme (SEQ ID NO: 9) is a multi-unit transporting or exporter enzyme. Methods of testing activity are well known in the art and described in: Stevenson et al., *J. Bacteriol.* 178(16):4885-4893, 1996; Glucksmann et al., *J. Bacteriol.* 175(21):7045-7055, 1993; and Smith et al., *Mol. Microbiol.* 4(11):1863-1869, 1990, which are herein incorporated by reference.

Finally, sequence analysis using Blast[™] searching indicates that the "U" enzyme (SEQ ID NO: 15) is a glycosyltransferase/exporter enzyme. Methods of

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testing glycosyltransferase/exporter activity are well known in the art and described in: Stevenson et al., *J. Bacteriol.* 178(16):4885-4893, 1996; Glucksmann et al., *J. Bacteriol.* 175(21):7045-7055, 1993; Smith et al., *Mol. Microbiol.* 4(11):1863-1869, 190; van Kranenburg et al., *J. Bacteriol.* 181(1):338-340, 1999; Kranenburg et al., *J. Bacteriol.* 181(11):6347-6353, 1999; Stingele et al., *J. Bacteriol.* 181(20):6354-6360, 1999.; Kolkman et al., *J. Bacteriol.* 178(13):3736-3741, 1996; Kolkman et al., *J. Biol. Chem.* 272(31):19502-19508, 1997; Breton et al., *Struct. Biol.* 9:563-571, 1999; and Griffiths et al., *J. Biol. Chem.* 273(19):11752-11757, 1998, which are herein incorporated by reference.

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Having illustrated and described the principles of the invention in multiple embodiments and examples, it should be apparent to those skilled in the art that the invention can be modified in arrangement and detail without departing from such principles. The invention encompasses all modifications coming within the spirit and scope of the following claims.

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CLAIMS

What is claimed is:

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- 1. An isolated bacterium having the characteristics of Lactococcus lactis subspecies cremoris Ropy 352, as deposited with the USDA-ARS-NCAUR-NRRL as deposit accession number NRRL B-30229.
- 10 2. A purified ropy polysaccharide wherein the polysaccharide has characteristics comprising:

Composition:

Glucose: range of 54% to 58%

Galactose: range of 42% to 46%

Charged:

Yes

Molecular weight:

range of 800,000 to 8,000,000

Phosphorous: Present in backbone or sidechain

Structure:

endpoints: galactose;

branchpoints: glucose

- 20 3. A purified ropy polysaccharide, isolated from Lactococcus lactis subspecies cremoris Ropy 352.
 - 4. The purified polysaccharide of claim 3 wherein the polysaccharide has the characteristics of:

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Composition:

Glucose: range of 54% to 58%

Galactose: range of 42% to 46%

Charged:

Yes

Molecular weight:

range of 800,000 to 8,000,000

Phosphorous: Present in backbone or sidechain

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Structure:

endpoints: galactose;

branchpoints: glucose

- 5. A method of thickening a liquid comprising adding to a liquid the purified polysaccharide of claim 2.
 - 6. The method of claim 5 wherein the liquid is a food.

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- 7. The method of claim 6 wherein the food is selected from the group consisting of milk, a milk-based liquid, a whey-based liquid, a soy-based liquid, and a fruit-juice.
- 10 8. A food product made by the method of claim 6.
 - 9. A method of thickening a liquid comprising adding to a liquid the purified polysaccharide of claim 3.
- 15 10. The method of claim 9 wherein the liquid is a food.
 - 11. The method of claim 10 wherein the food is selected from the group consisting of milk, a milk-based liquid, a whey-based liquid, a soy-based liquid, and a fruit-juice.

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- 12. A food product made by the method of claim 10.
- 13. A method of making a food product comprising addition of a culture of Ropy 352 to a food that is devoid of Ropy 352.

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- 14. The method of claim 10 wherein the food is selected from the group consisting of milk, a milk-based liquid, a whey-based liquid, a soy-based liquid, and a fruit-juice.
- 30 15. A food product made by the method of claim 13.

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16. An isolated plasmid of approximately 20 kb derived from *Lactococcus lactis* subspecies *cremoris* Ropy 352, wherein the plasmid, when expressed in the transformed lab strain of *Lactococcus* MG1363, expresses a ropy polysaccharide, wherein the polysaccharide has characteristics comprising:

Composition:

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Glucose: range of 54% to 58%

Galactose: range of 42% to 46%

Charged:

Yes

Molecular weight:

range of 800,000 to 8,000,000

Phosphorous: Present in backbone or sidechain

Structure:

endpoints: galactose;

branchpoints: glucose

17. A probe comprising a detectable label attached to a nucleic acid selected from the group consisting of:

a portion of the plasmid of claim 16, and the plasmid of claim 16.

- 18. A method of detecting a target nucleic acid comprising the steps of:
 contacting the target nucleic acid with the probe of claim 17 under
 conditions wherein the probe hybridizes with the target nucleic acid, and
 detecting the detectable label.
 - 19. A cell transformed with the plasmid of claim 16.
- 25. The cell of claim 19, wherein the cell is selected from the group consisting of: a bacterial cell, a yeast cell, a fungal cell, an animal cell and a plant cell.
- 21. A method of making a food product comprising addition of the cell of claim 16 to a food that is devoid of the plasmid of claim 16.
 - 22. A method for making a pharmaceutical product comprising:

combining an active ingredient and the purified ropy polysaccharide of claim 2.

23. A pharmaceutical product made by the method of claim 22.

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- A method of making a beauty care product, comprising adding the 24. purified ropy polysaccharide of claim 2.
 - 25. A beauty care product made by the method of claim 24.

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- 26. A method of making a coating agent, comprising adding the purified ropy polysaccharide of claim 2.
 - 27. A coating agent made by the method of claim 26.

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- 28. A purified protein, comprising an amino acid sequence selected from the group consisting of:
- (a) an amino acid sequence selected from the group consisting of SEQ ID NOS: 9, 10, 13, 14, and 16;

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- (b) an amino acid sequence that differs from those specified in (a) by one or more conservative amino acid substitutions; and
- (c) an amino acid sequence having at least 60% sequence identity to the sequences specified in (a).
- 29. An isolated nucleic acid molecule encoding a protein according to claim 28.
 - 30. An isolated nucleic acid molecule, comprising a nucleic acid sequence selected from the group consisting of:
- 30 (a) a nucleic acid sequence selected encoding an amino acid sequence selected from the group consisting of: SEQ ID NOS: 9, 10, 13, 14, and 15;

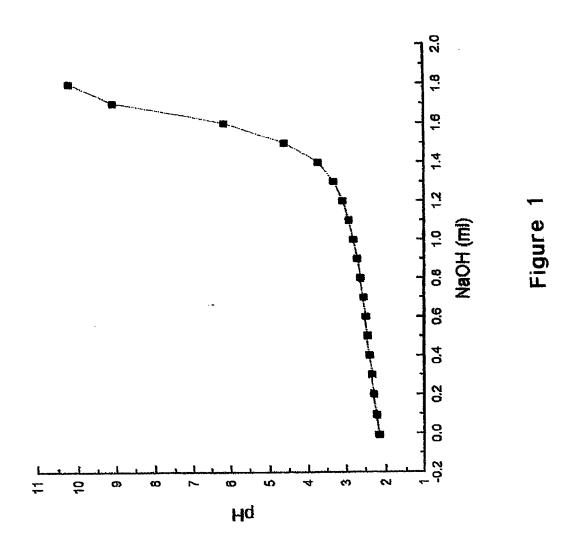
- 32 -

- (b) a nucleic acid sequence that shares at least 60% sequence identity with the nucleic acid sequences described in (a);
- (b) an nucleic acid sequence that comprises at least 15 consecutive nucleotides of the sequences shown in (b).

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- 31. A recombinant nucleic acid molecule comprising a promoter sequence operably linked to a nucleic acid sequence according to claim 30.
- 32. A cell transformed with a recombinant nucleic acid molecule according to claim 31.
 - 33. A transgenic bacteria comprising a recombinant nucleic acid according to claim 31.
- 15 34. A method of producing a protein, comprising:

 culturing a cell according to claim 32, wherein the cell expresses at least one protein from the recombinant nucleic acid; and isolating the protein.



Expression EPS352 for Necessary (2265 base pairs from the start of EpsM to the end of EpsN) Epsn) (EpsM and Two Genes O. Sequence

cagagagaaaattattaaaaagggaacttaattaagcttaaaattgggggagtataaaaootnoteminscagagaaaatttaattaattagtaccagtttataattcagaa ootnotemins ootnotemingtototottttaataaatttttooottgaattaattogaattttaacooootoatattttaaotogottttaaattagtoataatatoatggtoaaatattaagtott

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aaaaatotgggtotgotgtgaatactatttcaatgacaaatotttactaaccoaactaattattçaaattacgactacaacaatactcattaatgatatatacgtttt ttttagacccagacgacacttatgataaaagttactgtttagaaatgattgggttgattaataagtttaatgctgatgttgttatgagtaattactatatgcaaa F L D P D D T Y D K S Y C L E M I G L I N K F N A D V V M S N Y Y I C K

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Figure 2A

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Figure 3A

B. Alignments 1) Alignment of EpsM to EpsN

Figure

FYDQYKIRELGAVVGKIVMTTLASFPDSEKLYNELNPIRKKVFKDYISIEK-RHTKRIKMYVKMYVFSSYVGYKLYRLVKGKHWK VIKRSVIEKHDLLFSVGKKFEDNNFTPKVFYFSKNIVVIS---LRLYRYRKRSGSIMS----NRPEKFFSDDAIFVTYD---LLD -MIKLSIIIPIYNVEKYLSKCLNSILEQTYKEIEIILVNDGSTDNSKDIAVSYCERFPN--VFKYFEKDNGGLSSARNFGLEKIS IYNVEWLEKINIKFKEGLLYEDLNFFFKIVPHLTSISEVSTVKNSFVHYVQHKGTITSDNSLNILDIIKSYEDVFHYYNEKQIND MNPLISIIVPIYNVEKYIGSLVNSLLKQTNKNFEVIFIDDGSTDESMQILKEIMAGSEQEFSFKLLQQVNQGLSSARNIGILNAT GEYLFFLDSDDEIESNFVETILTSCYKYSQPDTLIFDYSSIDEFGNALDSNYGHGSIYRQKDLCTSEQILTALSKDEIPTTAWSF GDFVGFLDSDDYIDNDLYEIMIN----SLDSSIKIVECDFIWEYEN-----GKSVLDKTSEYNSIKDLMVNG--RVV---AWNK LYFDELEYKFSRNLMGAFLKRAIKIKDKRQRKIILDEFWNNVLSYYPNWKKNKYIKKLSKQNILLFFINKYTYKLFYLL-----LSENLISIIVPVYNSEKYLRAAIHSLLNQTYQNIEVILINDGSTDGSQELISSFQKKD-KRIKLYNTKNLGVSHARNYGIDRASG --MIKLSIIIPIYNVEKYLSKCLNSILEQTYKEIEIILVNDGSTDNSKDIAVSYCERFPNVFKYFEKDNGGLSSARNFGLEKISG EWLEKINIKFKEGLLY-EDLNFFFKIVPHLTSISEVSTVKNSFVHYVQHKGTITSDNSLNILDIIKSYEDVFHYYNEKQINDLYF SY IMFLDPDDTYDKSYCLEMIGLINKFNADVVMSNYYICKGKNIYPNVNNDLLECEGLLSRDKTMRSILSDTGFKGFVWTRIFRK DFVGFLDSDDYIDNDLYEIMINSL---D----SSIKIVECDFIWEYENGKSVLDK--TSEYNSIKDLMVNG--RVVAWNKIYNV nvin--nvkfnesinyledmlfnisivhnarilaytnkrhyfyloredsaskkfsksffkslnlirgkvdpefysgidsv--ify ** ******* * **** ** NLVGWLITERKSRENSQFIRRNIKN-MKSQVKFKTLKMENPIKNLILKLSYAFPLVGSCMIHMLSVFMKTKLYSKLMSMLRKG 2) Alingment of EpsM to EpsG (a Lactococcus lactis glycosyltransferase involved in a different EPS operon) DELEYKFSRNLMG---AFLKRAIKIKDKRQRKIILDEFWNNVLSYYPNWKKNKYIKKLSKQNILLFFINKYTY-KLFYLL--* .. *** ** * * * * * * * •• • • ** * ******************* .. * *: * **** ****** * ******** 3) Alingment of EpsN to EpsG EpsG EpsN EpsG EpsN EpsG EpsN EpsG EpsG EpsM EpsM EpsG EpsM EpsG

Organization of pEPS352

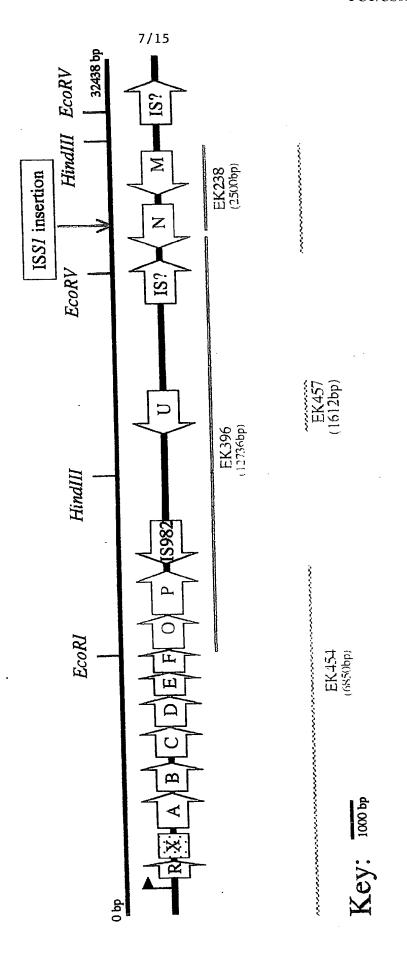


Figure 4

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Figure 5A

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Figure 5B

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Figure 5C

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Figure 5D

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caaccatttaatcgactcattcaaaaaattgaaggaacttgtacgcgatggtgaaatcgaagacgatgtattcatgcaaattgggtactcaacttatgaac GTTGGTAAATTAGCTGAGTATTTTTAACTACTTGAACATGCGCTACCACTTTAGCTTCTGCTACATAAGTACGTTTAACCCATGAGTTGAATACTTG ტ н α Σ Ŀ > Ω Δ ы ы ഗ œ ы Ω Н × Ø

GATTTATATGATTTACCCTTTCAAATAACCTATACTTTCTACATACTTACTTCGCTCATGCTAATAATGAGTACCGCCTGGTAGATGGAT P K Y T K W E K F I G Y E T M E R C M N E A S T I I T H G G P S T Y

CAACACTITITCCCTATGAGTAACTACAACTICTGTAAGAGCTTTTATAATAATCAAGGTTTTAAAGTCTATGGAATGTTTTTTTAC f v o f k gttgtgaaaaagggatactcattgattttgtgcgaagatgttgaagacattctcgaaaatattattagttccaaaatttcagataccttacaaaaaatg

Figure 5E

atttagtgttgtgacttaagtattttaataagtcacgactttaaatggtcgataaatattttcactcttctatactatggttttcattatgtgataacg tggttcggagggcaacctttaccagaatctgcgctaaaatgtattgaaagttggagaaggttttgtccagattatgaaataaaacaatggtctgagaaaa ρι н ជា Σ >-× 回 പ ပ ഗ 幺 н Œ H Ø >1 Н 띠 ď ഗ ĪΣ ы × Н

ipaatcacaacactgaattcataaaattattcagtgctgaaatttaccagctatttataaaaagtgagaagatatgataccaaaagtaatacactattgc

accaagcctcccgttggaaatggtcttagacgcgattttacataactttcaacctcttccaaaacaggtctaatactttattttttaccagactctttt ſτι α αú 3 တ ပ × 4 ഗ 回 Q, н Д Ø

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TCCGCCATATATAGAACTIGTGCCTACATCTCGAATATTTAGAGAACTACTTAACGACATATTATCAAATATAAATCCTTACCTTTCTCGACCATCTCAT G G I Y L D T D V E L I K S L D E L L Y N S L Y L G M E R A G R V aggcggtatatatetteacacggatgtagagcttataaaatctcttgatgaattgctgtataatagtttatatttggaatggaaagagctggtagagta

aatacgggtttaggggtttggagctgaagtaaatcatccaattgtgagagctaatttagaattgtatactaatattcctttttcaggcaatgataatata ttatgeccaaateccaaacetegaettagtaggttaacaetecgattaaatettaacatataaatetaatgattataaggaaaaagtecgttaetattatatt ဟ Д H Z H × ы 凶 н z 4 œ > н Д Ħ z > M Ø ശ CTTGTGTGACCTATACGACGAATCTTTTGAAAAATATGGTCTAAAAACAACAAGAAATTCAACATATAGATAACGCAATAATTTTACCTACTGAATA GAACACACTGGATATGCTGCTTAGAAAACTTTTTATACCAGATTTTTTGTTGTTGTTAAGTTGTATTTGCGTTATTAAAATGGATGATGATTAT C V T Y T T N L L K K Y G L K N N N E I Q H I D N A I I L P T E Y

 ĸ × × Œ ы [zı Ω Д G 3 × œ Ø CATGACAAGAGAGAGAGTGATTATGCCTTATGTGTGGTGGTAATTTTAGAATATAATAATAATACAGGATTAATTGCGTCTTCAGCATACTTTTAGCATG GTACTGTTCTCTCTAATAACGGAATACACATTAAAATCTTATAAATTTATGTCCTAATTAACGCAGAAGTCGTATGAGAAAATCGTAC M T R E M R V I A L C V V I L E Y L N N T G L I A S S A Y S F S M

> н н × G × ፈ × × ပ ĹΣι ы >-1 ы

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Σ Ω ល ഗ 노 z н S Σ Н × н > Н E⊣ ы н Бч ပ > ഗ GCTTTTADAATCGTATGAGTTTAGGATTTATCCAACCGAATTTTGCAATGATGAGCTTTTTAGGTATAGCGATAGCCTTATTATATTTGAGTACTGAAA cgaaaatatttagcatactcaaatcctaaataggttggattaaaacgttactactcgaaaaatccatatcgctatcggaataataaactcatgacttt ď ď ග ы Бų ഗ M ø Ŀ z Ч O н [ī1 ტ Ы . V) Σ ĸ >ტ တ ₽ æ ഗ ø E Ŀı ы (L) נבו Н н æ 1-1 ᄄ

ataaaataaacaatcattttttttttttttcgttcaaagtttaaaacttttttcctcgtaatgtcaaaatggtgatgaaaaattagtagaaaat tattttatttigttagtaaaaaaactaaaagcaagtttcaaattttgaaaaaaggagcattacagttttaccactacttttaatcatcatcttatt Ц H ч Н D. ы > E н ഗ œ × 凶 ĮΉ z ഗ > Ø × × ᇊ × × ഗ S >1 н 团 O. **>**+ H ø ы **~** ശ ഗ ы ы ഗ z Н × O z

ATCCCTIATTACTACAATTTTATGTTACAATGTCGTATAGAAGGTTCAAACGATCGTTTTCCTTAAAAAAATGTAACAAAAATAAACATTGAAA I G N N D V K N T M L D T A Y L Q S L L A K G I L F T L F U F V T F

CTTITICATATTTTTTTAAGAGAAAAACACAAACTAGGTTGCAAAGTTTAGTAATTATGATGTATTTTTAATTGCATTTACAGAAACATCATTTTTT gaaaagtataaaaaagaattctctttttgtgtttgatccaacgtttcaaatcattaatactacataaaaaattaacgtaaatgtctttgtagtaaaaa ٤٦ بعرا K н ᆸ ជរ × Σ Σ Н > н ഗ Ø н œ ₽ 0 E-I × œ ×

TCCAAACATTAAAATAAAGGTCATAACTACCATTATTACCTAGTCTTTCTCCGATTATTTCATTATCTTTCCACCGTATCACTCATAATTATTTTGTCT aggittigtaattttatttccagtattgatggtaataatggatcagaaggggctaataaaagtaatagaaaaggtggcatagtggtattaataaaacaga × 团 н > × z ď ы × O₁ Ω Σ н > Σ ы > Д Īщ

GATTGAGGAATACAAAGTATCCGTTATAGTTCCTGTTTACAATGTAGAGG CTAACTCCTTCTGTTTCATAGGCAATATCAAGGACAAATGTTACATCTCC

Figure 5G

15/15

Sequence of EpsU (start and stop codons are underlined) 1612bp total here but 1412 from start codon to stop codon

GGTGGACAGGAGGACACAATTTTTAATCCTTCCTGTTATATAGTTTTTGTTTAATATTTTTTCGGGAGGGTT ATTA<u>ATG</u>CAAATCGCAAAAAATTATCTTTATAATGCAATATATCAGGTCTTTATAATAATTGTGCCATTAC TTACCATTCCTTATTTGTCAAGAATTTTGGGCCCTTCAGGTATTGGAATTAACTCATATACCAATTCTATT GTTCAGTATTTTGTTTTATTTGGTAGTATAGGAGTCGGTTTGTATGGGAATCGTCAGATTGCCTTTGTTAG GCTATAGTTGCAGCTGCATTTGATATCTCTTGGTTTTTTTATGGGAATTGAAAATTTTAAAGTAACTGTATT AAGAAATTTTATAGTTAAGTTACTTGCTCTATTCAGTATTTTCCTATTTGTCAAATCTTACAATGATTTGA **ATATATATATATTGATAACAGTTTTATCTACATTAATTGGTAATTTAACTTTTTTCCCAAGTTTACACAGA** TATCTCGTAAAGGTTAACTATCGTGAATTAAGGCCAATAAAGCATTTAAAGCAATCTTTAGTCATGTTTAT CCCACAAATTGCTGTCCAAATTTATTGGGTTTTGAATAAAACGATGTTAGGTTCATTGGATTCTGTCACGA GCTCCGGCTTTTTTGATCAGTCTGATAAAATAGTTAAACTGGTTTTGGCTATTGCTACTGCAACAGGTACT GTCATGTTGCCACGTGTTGCAAATGCCTTTGCACATAGAGAGTATAGTAAAATTAAGGAATACATGTACGC AGGTTTTTCTTTTGTGTCGCCAATTTCGATTCCTATGATGTTTGGTCTGATAGCTATTACTCCTAAATTCG TGCCACTTTTTTTTACATCTCAATTTAGTGATGTTATTCCTGTGTTAATGATCGAGTCAATCGCAATTATT TTTATAGCTTGGAGCAACGCAATAGGTACTCAATATCTTTTACCAACTAATCAAAATAAGTCATATACAGT GTCGGTGATCATTGGAGCGATAGTCAATTTAATGTTAAATATTCCACTGATTATATATCTAGGTACTGTTG GTGCATCAATTGCAACTGTAATTTCTGAAATGTCTGTAACTGTGTATCAACTTTTTATAATTCATAAACAG CTTAATTTGCATACACTGTTTGCGGATTTATCTAAGTATTTAATTGCAGGATTAGTGATGTTTCTAATTGT AGGTATGGATTTAGGTACCTGCCTTATTGAAAATAACGGTGAGTCAATGGTATTGGGCATATTTGACGCTC ACCTTCAATTTGTTTTGGTCGACTTGATTGTAGCACAGGACAATATGTCT

SEQUENCE LISTING

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cagaaaa	att aag	gttttt	c tt	acag	gaag	t ta	ataa	aaaa	aggi	gatt	ata		atg Met 1	176
	tta tt Leu Ph													224
	gca aa Ala As 20			_		_	_				_			272
ttg aat Leu Asr 35	Asn Ty	_	_			_					_			320
gga cta Gly Leu 50	_	•								_				368
att gat Ile Asp			_	_		_								416
act caa Thr Glr		Lys		_	_				_					464
ttt tta Phe Leu	-					taa	caat	caata	aaa t	ttaq	gga	gt		511
					100									
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ggc gaa Gly Glu	aca ag Thr Se act aa	As gaa Glu cct	caa Gln gat	tta Leu 125 gtt	tt to ne Tr 11 ttg Leu	gct Ala	gaa Glu tat	aaa Lys gaa	gtt Val 130 gct	caa Gln	le Th 15 aat Asn ctt	caa Gln ttt	yr Asn gta Val aat	
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ggc gaa Gly Glu 120 ttg gcg Leu Ala 135	aca ag Thr Se act aa Thr As caa aa Gln As	As gaa Glu cct Pro att Ile 155	caa Gln gat Asp 140 gaa Glu	tta Leu 125 gtt Val gca Ala	ttg ttg Leu gtt Val aca Thr	gct Ala tta Leu gcc Ala	gaa Glu tat Tyr tca Ser 160 gca	aaa Lys gaa Glu 145 tgg Trp	gtt Val 130 gct Ala act Thr	caa Gln cca Pro agt Ser	le The left of the	caa Gln ttt Phe gag Glu 165 caa	gta Val aat Asn 150 caa Gln	611 659

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agt tac co Ser Tyr Pr 215	-	_				-	_
gat gga gt Asp Gly Va	_		-		_		
gat tat at Asp Tyr Il			_		taagtt a	ıtaaataad	ca 997
attattaaat	attggagaa	-		_	g gaa cag n Glu Gln	_	a Asp
tta aga gg Leu Arg Gl							
ttt agt go Phe Ser Al 28	a Leu Ile	Val Thr I					
ata gcc tc Ile Ala Se 300	_	-	-		_	-	
cca aat to Pro Asn Se 315							
att caa at Ile Gln Me			Asn Gln				
tta gat aa Leu Asp Ly	_	_			Asp Gly		
aaa caa gt Lys Gln Va 36	l Thr Val	Ála Asn G		-	_	_	
act gtt aa Thr Val Ly 380				Ala Gln	_		
act gct aa Thr Ala Ly 395							
aac gtt aa Asn Val As		_	la Lys .	_			-

			a tot gtt ata e Ser Val Ile 5		
	Ala Ile Ala		g gaa tta ttt s Glu Leu Phe		
			g ggg ctc acg n Gly Leu Thr 470	Val Leu Gl	
		Ser Asp Phe	aat aag aat Asn Lys Asn 485		
	3 3 3		ccg cct agc Pro Pro Ser 500		ı Val
	tca aaa agg Ser Lys Arg 510			gg atg gct a Met Ala I	
	_	_	att att acc Ile Ile Thr 530		n Pro
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	-	_	att gaa gat Ile Glu Asp	-	
			att acc tct Ile Thr Ser		
			agt gca atg Ser Ala Met 660		
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665 670 675 680	.)
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gtt ttg cat ggg gta aac tct tct gag tca cca tcg tat tac tac cac Val Leu His Gly Val Asn Ser Ser Glu Ser Pro Ser Tyr Tyr His 730 735 740	
gga gta gag taa ttggaataaa cttgaatcaa ataaaagaca gaaatttgta Gly Val Glu 745	2544
gaagaggaga gcaaatgatt gatattcatt gccatatttt actggagcta aaacttct	gg 2604
agatactttg aca atg ctg aaa tca gca att gat gaa ggg ata aca acc Met Leu Lys Ser Ala Ile Asp Glu Gly Ile Thr Thr 750 755 760	2653
atc act gcc act cct cat cat aat cct caa ttt aat aa	
ctt att ttg aag aaa gtt aag gaa gtt caa aat atc att gac gag cat Leu Ile Leu Lys Lys Val Lys Glu Val Gln Asn Ile Ile Asp Glu His 780 785 790	
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795 800 805	
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aaa atg ata Lys Met Ile 905	-		_	_	-			3133
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gaa gat agt Glu Asp Sei		_	_	Arg Met			-	3229
gag tca gto Glu Ser Val 955	lle Leu	-	-		_			3277
atc aaa aca Ile Lys Thi 970	_				aaggatta	aaa aggag	gtaaat	3330
a atg gaa t Met Glu F 980				er Pro Gi				3379
tta gta gaa Leu Val Glu			Ser Tyr					3427
gca att gat Ala Ile Asp					Val Leu			3475
aca act ac								
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Ala Ala Ala 1030 caa ggg cca Gln Gly Pro 1045 att ttt tat Ile Phe Tyr	atg ttc Met Phe att ttg	tat aaa Tyr Lys 1050 aaa ttt Lys Phe 1065 cca gat	Pro Tyr 1035 caa aaa Gln Lys aga aca Arg Thr gtt aaa Val Lys	cgc tat Arg Tyr atg att Met Ile 1070 gct gct	Ser Ser 1040 ggt aaa Gly Lys .055 ctt aat Leu Asn	Lys Lys aat ggt Asn Gly gcc gag Ala Glu 1 gcc aac	Asp aaa : Lys cag : Gln 075	3571
Ala Ala Ala 1030 caa ggg cca Gln Gly Pro 1045 att ttt tat Ile Phe Tyr 1060 tat cta gaa	atg ttc Met Phe att ttg Ile Leu ctt aat Leu Asn 1080	Tyr Ile tat aaa Tyr Lys 1050 aaa ttt Lys Phe 1065 cca gat Pro Asp	Pro Tyr 1035 caa aaa Gln Lys aga aca Arg Thr gtt aaa Val Lys cgg gta	cgc tat Arg Tyr atg att Met Ile 1070 gct gct Ala Ala 1085 acg aag	Ser Ser 1040 ggt aaa Gly Lys .055 ctt aat Leu Asn tac cat Tyr His att ggc Ile Gly	Lys Lys aat ggt Asn Gly gcc gag Ala Glu 1 gcc aac Ala Asn 1090 tca ttt	Asp aaa : Lys cag : Gln 075 ggc : Gly ata :	3571 3619
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caa cga gca gat tta gaa ctc tat tat ctc cag tac cat agc acc aaa 39 Gln Arg Ala Asp Leu Glu Leu Tyr Tyr Leu Gln Tyr His Ser Thr Lys 1175 1180 1185	955
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gat aga ttt tgg gtc aca ttt gat aaa aca gat gca aaa tct ata ttg 41 Asp Arg Phe Trp Val Thr Phe Asp Lys Thr Asp Ala Lys Ser Ile Leu 1235 1240 1245 1250	47
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gct ttt gtc acg gat gtt gca agg ctc gat ata att tgg aat gaa ggc 5204 Ala Phe Val Thr Asp Val Ala Arg Leu Asp Ile Ile Trp Asn Glu Gly 1420 1425 1430 1435	4
ggt ata tat ctt gac acg gat gta gag ctt ata aaa tct ctt gat gaa 5252 Gly Ile Tyr Leu Asp Thr Asp Val Glu Leu Ile Lys Ser Leu Asp Glu 1440 1445 1450	<u>3</u> .
ttg ctg tat aat agt tta tat tta gga atg gaa aga gct ggt aga gta 5300 Leu Leu Tyr Asn Ser Leu Tyr Leu Gly Met Glu Arg Ala Gly Arg Val 1455 1460 1465)
aat acg ggt tta ggg ttt gga gct gaa gta aat cat cca att gtg aga 5348 Asn Thr Gly Leu Gly Phe Gly Ala Glu Val Asn His Pro Ile Val Arg 1470 1475 1480	3
gct aat tta gaa ttg tat act aat att cct ttt tca ggc aat gat aat 5396 Ala Asn Leu Glu Leu Tyr Thr Asn Ile Pro Phe Ser Gly Asn Asp Asn 1485 1490 1495	5
ata act tgt gtg acc tat acg acg aat ctt ttg aaa aaa tat ggt cta 11e Thr Cys Val Thr Tyr Thr Asn Leu Leu Lys Lys Tyr Gly Leu 1500 1505 1510 1515	ļ
aaa aac aac aat gaa att caa cat ata gat aac gca ata att tta cct Lys Asn Asn Glu Ile Gln His Ile Asp Asn Ala Ile Ile Leu Pro 1520 1525 1530	?
act gaa tat tta tgt cct cta agt ttt gaa aca aat cga tta aaa ata 5540 Thr Glu Tyr Leu Cys Pro Leu Ser Phe Glu Thr Asn Arg Leu Lys Ile 1535 1540 1545)

acg gaa aat act tac tcc atc cat cac tat gat atg agt tgg aaa gat 5 Thr Glu Asn Thr Tyr Ser Ile His His Tyr Asp Met Ser Trp Lys Asp 1550 1560	588
aag aga gat aaa ttt tta aga ctt aaa ata caa ctt aga aaa tgg gta 5 Lys Arg Asp Lys Phe Leu Arg Leu Lys Ile Gln Leu Arg Lys Trp Val 1565 1570 1575	636
ggt gat gat ttt tat gaa aaa gtt att aaa aga att gga aaa taa t	686
atg aat aaa ata acc atg aca aga gag atg aga gtt att gcc tta tgt 5°Met Asn Lys Ile Thr Met Thr Arg Glu Met Arg Val Ile Ala Leu Cys 1595 1600 1605 1610	734
gtc gta att tta gaa tat tta aat aca gga tta att gcg tct tca 5° Val Val Ile Leu Glu Tyr Leu Asn Asn Thr Gly Leu Ile Ala Ser Ser 1615 1620 1625	782
gca tac tct ttt agc atg gcg agt aca atc ctc tta tcc tat atc tta 58 Ala Tyr Ser Phe Ser Met Ala Ser Thr Ile Leu Leu Ser Tyr Ile Leu 1630 1635 1640	830
ttc tgt aaa aaa aga aaa gga ttt tct tta aag gag att att gta cta 58 Phe Cys Lys Lys Arg Lys Gly Phe Ser Leu Lys Glu Ile Ile Val Leu 1645 1650 1655	878
cta att cca ttt att ttt gta gtt tta aat cgt gat cct agt aat ttc 59 Leu Ile Pro Phe Ile Phe Val Val Leu Asn Arg Asp Pro Ser Asn Phe 1660 1665 1670	926
agt tta ggg tta atg tgg ata ctc tat ttt atg tta agt aag tcg gaa 59 Ser Leu Gly Leu Met Trp Ile Leu Tyr Phe Met Leu Ser Lys Ser Glu 1675 1680 1685 1690	974
ata gat tta aaa aaa gtg atg aaa aca ttt ttt gtt acc tct agt gtt 60 Ile Asp Leu Lys Lys Val Met Lys Thr Phe Phe Val Thr Ser Ser Val 1695 1700 1705	022
tgt ttt att ttg aca ata gta ctt tat tta ata atg tct ctt aat aaa 60 Cys Phe Ile Leu Thr Ile Val Leu Tyr Leu Ile Met Ser Leu Asn Lys 1710 1715 1720	070
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agt tta gga ttt atc caa ccg aat ttt gca atg atg agc ttt tta ggt 61 Ser Leu Gly Phe Ile Gln Pro Asn Phe Ala Met Met Ser Phe Leu Gly 1740 1745 1750	166
ata gcg ata gcc tta tta tat ttg agt act gaa aga caa aga ata act Ile Ala Ile Ala Leu Leu Tyr Leu Ser Thr Glu Arg Gln Arg Ile Thr 1755 1760 1765 1770	214
ata att ttt att gcc att gta act ttt att ata ttt tac ttt act caa 62 Ile Ile Phe Ile Ala Ile Val Thr Phe Ile Ile Phe Tyr Phe Thr Gln 1775 1780 1785	262

tca aga act tca gga tat atc tta ttt ttt att ttg agt att tta ttt Ser Arg Thr Ser Gly Tyr Ile Leu Phe Phe Ile Leu Ser Ile Leu Phe 1790 1795 1800	6310
gtt agt agt aaa aaa act aaa aag caa gtt tca aat ttt gaa aaa agg Val Ser Ser Lys Lys Thr Lys Lys Gln Val Ser Asn Phe Glu Lys Arg 1805 1810 1815	6358
agc att aca gtt tta cca cta ctt ctt tta atc atc tct tat tcg ttg Ser Ile Thr Val Leu Pro Leu Leu Leu Ile Ile Ser Tyr Ser Leu 1820 1825 1830	6406
tta aag tta cct att aat caa tac atc aat agc ttg ctt tct ggt cgt Leu Lys Leu Pro Ile Asn Gln Tyr Ile Asn Ser Leu Leu Ser Gly Arg 1835 1840 1845 1850	6454
ctg gcg ctt tat caa gag att tat tct aca ttt ggt ata cat ttg ata Leu Ala Leu Tyr Gln Glu Ile Tyr Ser Thr Phe Gly Ile His Leu Ile 1855 1860 1865	6502
ggg aat aat gat gtt aaa aat aca atg tta gat aca gca tat ctt caa Gly Asn Asn Asp Val Lys Asn Thr Met Leu Asp Thr Ala Tyr Leu Gln 1870 1875 1880	6550
agt ttg cta gca aaa gga att ttg ttt aca ttg ttt tta ttt gta act Ser Leu Leu Ala Lys Gly Ile Leu Phe Thr Leu Phe Leu Phe Val Thr 1885 1890 1895	6598
ttc ttt ttc ata ttt ttt ctt aag aga aaa aca caa act agg ttg caa Phe Phe Phe Ile Phe Phe Leu Lys Arg Lys Thr Gln Thr Arg Leu Gln 1900 1905 1910	6646
agt tta gta att atg atg tat ttt tta att gca ttt aca gaa aca tca Ser Leu Val Ile Met Met Tyr Phe Leu Ile Ala Phe Thr Glu Thr Ser 1915 1920 1925 1930	6694
ttt ttt agg ttt gta att tta ttt cca gta ttg atg gta ata atg gat Phe Phe Arg Phe Val Ile Leu Phe Pro Val Leu Met Val Ile Met Asp 1935 1940 1945	6742.
cag aaa gag gct aat aaa gta ata gaa aag gtg gca tag tgagtattaa Gln Lys Glu Ala Asn Lys Val Ile Glu Lys Val Ala 1950 1955	6791
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Gly Lys Ser Ala Asn Gln Ile Glu Arg Glu Leu Gly Tyr Pro Arg Asn 20 25 30 Ser Leu Asn Asn Tyr Lys Leu Cly Cly Cly Pro Ser Cly Thr Arg Leu	
Ser Leu Asn Asn Tyr Lys Leu Gly Gly Glu Pro Ser Gly Thr Arg Leu 35 40 45	
Ile Gly Leu Ser Glu Tyr Phe Asn Val Ser Pro Lys Tyr Leu Met Gly 50 55 60	

 Ile Ile Asp Glu Pro Asn Asp Ser Ser Ala Ile Asn Leu Phe Lys Thr
 65
 70
 75
 80

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 90
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 105

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145
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                                   155
Lys Ala Lys Ala Gln Thr Thr Pro Ile Ser Pro Lys Pro Lys Leu Tyr
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              165
Leu Ala Ile Ser Val Ile Ala Gly Leu Val Leu Gly Leu Ala Ile Ala
                            185
Leu Leu Lys Glu Leu Phe Asp Asn Lys Ile Asn Lys Glu Glu Asp Ile
                         200
Glu Ala Leu Gly Leu Thr Val Leu Gly Val Thr Ser Tyr Ala Gln Met
                     215
                                      220
Ser Asp Phe Asn Lys Asn Thr Asn Lys Asn Gly Thr Gln Ser Gly Thr
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Lys Ser Ser Pro Pro Ser Asp His Glu Val Asn Arg Ser Ser Lys Arg
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Asn Lys Arg
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<210> 5 <211> 231 <212> PRT <213> Lactococcus lactis

<400> 5

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<210> 6 <211> 230 <212> PRT <213> Lactococcus lactis

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<210> 7

<211> 228

<212> PRT

<213> Lactococcus lactis

<400> 7

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Gln Arg Ala Asp Leu Glu Leu Tyr Tyr Leu Gln Tyr His Ser Thr Lys 200 Asn Asp Ile Lys Leu Leu Val Leu Thr Ile Val Gln Ser Ile Asn Gly 210 215 Ser Asp Ala Tyr 225 <210> 8 <211> 149 <212> PRT <213> Lactococcus lactis <400> 8 Met Lys Ile Ala Leu Val Gly Ser Ser Gly Gly His Leu Thr His Leu 10 Tyr Leu Leu Lys Lys Phe Trp Glu Asn Glu Asp Arg Phe Trp Val Thr 20 25 Phe Asp Lys Thr Asp Ala Lys Ser Ile Leu Lys Glu Glu Arg Phe Tyr 40 Pro Cys Tyr Tyr Pro Thr Asn Arg Asn Val Lys Asn Thr Ile Lys Asn 55 Thr Ile Leu Ala Phe Lys Ile Leu Arg Lys Glu Lys Pro Asp Leu Ile 75 70 . Ile Ser Ser Gly Ala Ala Val Ala Val Pro Phe Phe Trp Leu Gly Lys 90 85 Leu Phe Gly Ala Lys Thr Val Tyr Ile Glu Ile Phe Asp Arg Ile Asp 105 100 Lys Pro Thr Leu Thr Gly Lys Leu Val Tyr Pro Val Thr Asp Lys Phe 125 120 Ile Val Gln Trp Glu Glu Leu Lys Lys Val Tyr Pro Lys Ala Ile Asn 135 Leu Gly Gly Ile Phe <210> 9 <211> 235 <212> PRT <213> Lactococcus lactis <400> 9 Met Ile Pro Lys Val Ile His Tyr Cys Trp Phe Gly Gly Gln Pro Leu Pro Glu Ser Ala Leu Lys Cys Ile Glu Ser Trp Arg Phe Cys Pro Asp Tyr Glu Ile Lys Gln Trp Ser Glu Lys Asn Tyr Asp Val Asn Lys Ile Gln Tyr Ile Lys Glu Ala Tyr Gln Glu Lys Lys Phe Ala Phe Val Thr Asp Val Ala Arg Leu Asp Ile Ile Trp Asn Glu Gly Gly Ile Tyr Leu Asp Thr Asp Val Glu Leu Ile Lys Ser Leu Asp Glu Leu Leu Tyr 90 Asn Ser Leu Tyr Leu Gly Met Glu Arg Ala Gly Arg Val Asn Thr Gly 100 105 Leu Gly Phe Gly Ala Glu Val Asn His Pro Ile Val Arg Ala Asn Leu 115 120 Glu Leu Tyr Thr Asn Ile Pro Phe Ser Gly Asn Asp Asn Ile Thr Cys 135 140 Val Thr Tyr Thr Thr Asn Leu Leu Lys Lys Tyr Gly Leu Lys Asn Asn

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<213> Lactococcus lactis

325

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330

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<220>

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60

108

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			gtt Val													204
			agc Ser					_	_		-					252
			aat Asn													300
_	_	_	ggt Gly	_			-			-		_	_			348
			tac Tyr 100													396
_	_	_	gtt Val		_					_						444
			gtt Val			-			_	_	_					492
	-		aca Thr	_						-						540
			aca Thr													588
			agc Ser 180													636
	-		aat Asn	_	_			_					_			684
			caa Gln													732
			tcc Ser													780
	_		att Ile	_		_						-				828
			aga Arg 260	_	_		-		_					-		876
att	aaa	aat	atg	aaa	tcc	caa	gtt	aag	ttt	aaa	acg	ctt	aaa	atg	gaa	924

Ile Lys Asn Met Lys Ser Gln Val Lys Phe Lys Thr Leu Lys Met Glu 275 280 285	
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cta ttg aaa caa acg aac aag aat ttt gag gtt att ttt att gat gac Leu Leu Lys Gln Thr Asn Lys Asn Phe Glu Val Ile Phe Ile Asp Asp 360 365 370	1455
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	470					475					480					
	ata Ile			_	_			_	-					_		1839
	ttt Phe				_				-			_				1887
	gtt Val					_	_					-		_		1935
	ttt Phe	_				-	_				_	_		_		1983
	aaa Lys 550	-					_	_		_	_	_				2031
_	gac Asp	_	-			_			_			_			_	2079
_	tat Tyr				_	_		-	_	_				_	_	2127
	aca Thr		-				_	-			_			_		2175
	cca Pro		_			-			_					_		2223
_	cat His 630						_		-		_		-			2271
	tat Tyr	-						_	_	-						2319
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<211> 332

<212> PRT

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- <212> PRT
- <213> Lactoccocus Lactis

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- Ile Leu Lys Glu Ile Met Ala Gly Ser Glu Gln Glu Phe Ser Phe Lys 50 55 60
- Leu Leu Gln Gln Val Asn Gln Gly Leu Ser Ser Ala Arg Asn Ile Gly 65 70 75 80
- Ile Leu Asn Ala Thr Gly Glu Tyr Ile Phe Phe Leu Asp Ser Asp Asp 85 90 95
- Glu Ile Glu Ser Asn Phe Val Glu Thr Ile Leu Thr Ser Cys Tyr Lys
 100 105 110
- Tyr Ser Gln Pro Asp Thr Leu Ile Phe Asp Tyr Ser Ser Ile Asp Glu
 115 120 125
- Phe Gly Asn Ala Leu Asp Ser Asn Tyr Gly His Gly Ser Ile Tyr Arg 130 135 140
- Gln Lys Asp Leu Cys Thr Ser Glu Gln Ile Leu Thr Ala Leu Ser Lys 145 150 155 160
- Asp Glu Ile Pro Thr Thr Ala Trp Ser Phe Val Thr Lys Arg Ser Val 165 . 170 . 175
- Ile Glu Lys His Asp Leu Leu Phe Ser Val Gly Lys Lys Phe Glu Asp 180 185 190
- Asn Asn Phe Thr Pro Lys Val Phe Tyr Phe Ser Lys Asn Ile Val Val 195 200 205
- Ile Ser Leu Arg Leu Tyr Arg Tyr Arg Lys Arg Ser Gly Ser Ile Met 210 215 220
- Ser Asn Arg Pro Glu Lys Phe Phe Ser Asp Asp Ala Ile Phe Val Thr 225 230 235 240
- Tyr Asp Leu Leu Asp Phe Tyr Asp Gln Tyr Lys Ile Arg Glu Leu Gly
 245 250 255
- Ala Val Val Gly Lys Ile Val Met Thr Thr Leu Ala Ser Phe Pro Asp 260 265 270
- Ser Lys Lys Leu Tyr Asn Glu Leu Asn Pro Ile Arg Lys Lys Val Phe \$275\$ 280 285

Lys Asp Tyr Ile Ser Ile Glu Lys Arq His Thr Lys Arq Ile Lys Met

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	_				aat Asn	-	_					_			_	591
					ggt Gly										_	639
		-	_	-	aac Asn		-	_								687
_				_	atg Met 210						_	-				735
	-	_			acg Thr	_				_	_		_	_	_	783
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	-		_		ggt Gly		_	_	_		_	_	_		-	879
	_		_		tat Tyr	_			_	-		_		-		927
				_	gca Ala 290		_			_	_			_		975
_					ttc Phe										-	1023
-	-				tta Leu	-										1071
_		_		_	ata Ile											1119
	_				gtg Val	_							_			1167
_					ctg Leu 370							-				1215
			-		tct Ser											1263

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gtg ggc ata att att tat gtt gtt tta tta ata ttt tta aag gca gaa Val Gly Ile Ile Ile Tyr Val Val Leu Leu Ile Phe Leu Lys Ala Glu 455 460	1455
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Gly Pro Ser Gly Ile Gly Ile Asn Ser Tyr Thr Asn Ser Ile Val Gln 35 40 45	
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Tyr Phe Val Leu Phe Gly Ser Ile Gly Val Gly Leu Tyr Gly Asn Arg	
Tyr Phe Val Leu Phe Gly Ser Ile Gly Val Gly Leu Tyr Gly Asn Arg 50 55 60 CGln Ile Ala Phe Val Arg Asp Asn Gln Val Lys Met Ser Lys Val Phe	
Tyr Phe Val Leu Phe Gly Ser Ile Gly Val Gly Leu Tyr Gly Asn Arg Gln Ile Ala Phe Val Arg Asp Asn Gln Val Lys Met Ser Lys Val Phe 65 Tyr Glu Ile Phe Ile Leu Arg Leu Phe Thr Ile Cys Leu Ala Tyr Phe	
Tyr Phe Val Leu Phe Gly Ser Ile Gly Val Gly Leu Tyr Gly Asn Arg Gln Ile Ala Phe Val Arg Asp Asn Gln Val Lys Met Ser Lys Val Phe 65 Tyr Glu Ile Phe Ile Leu Arg Leu Phe Thr Ile Cys Leu Ala Tyr Phe 85 90 Ser Val Gly Gln Tyr His Ala Tyr Tyr	
Tyr Phe Val Leu Phe Gly Ser Ile Gly Val Gly Leu Tyr Gly Asn Arg Gln Ile Ala Phe Val Arg Asp Asn Gln Val Lys Met Ser Lys Val Phe 70 Phe 80 Tyr Glu Ile Phe Ile Leu Arg Leu Phe Thr Ile Cys Leu Ala Tyr Phe 85 Phe 90 Fle Val Ala Tyr Tyr 100 Leu Ser Gln Ser Ile Ala Ile Val Ala Ala Ala Phe Asp Ile Ser Trp	

Tyr Asn Asp Leu Asn Ile Tyr Ile Leu Ile Thr Val Leu Ser Thr Leu 170 Ile Gly Asn Leu Thr Phe Phe Pro Ser Leu His Arg Tyr Leu Val Lys 185 Val Asn Tyr Arg Glu Leu Arg Pro Ile Lys His Leu Lys Gln Ser Leu Val Met Phe Ile Pro Gln Ile Ala Val Gln Ile Tyr Trp Val Leu Asn 215 Lys Thr Met Leu Gly Ser Leu Asp Ser Val Thr Ser Ser Gly Phe Phe 235 Asp Gln Ser Asp Lys Ile Val Lys Leu Val Leu Ala Ile Ala Thr Ala Thr Gly Thr Val Met Leu Pro Arg Val Ala Asn Ala Phe Ala His Arg 265 Glu Tyr Ser Lys Ile Lys Glu Tyr Met Tyr Ala Gly Phe Ser Phe Val Ser Ala Ile Ser Ile Pro Met Met Phe Gly Leu Ile Ala Ile Thr Pro Lys Phe Val Pro Leu Phe Phe Thr Ser Gln Phe Ser Asp Val Ile Pro 315 Val Leu Met Ile Glu Ser Ile Ala Ile Ile Phe Ile Ala Trp Ser Asn Ala Ile Gly Thr Gln Tyr Leu Leu Pro Thr Asn Gln Asn Lys Ser Tyr Thr Val Ser Val Ile Ile Gly Ala Ile Val Asn Leu Met Leu Asn Ile Pro Leu Ile Ile Tyr Leu Gly Thr Val Gly Ala Ser Ile Ala Thr Val Ile Ser Glu Met Ser Val Thr Val Tyr Gln Leu Phe Ile Ile His Lys Gln Leu Asn Leu His Thr Leu Phe Ala Asp Leu Ser Lys Tyr Leu Ile 410 Ala Gly Leu Val Met Phe Leu Ile Val Phe Lys Ile Ser Leu Leu Thr 425 Pro Thr Ser Trp Ile Phe Ile Leu Leu Glu Ile Thr Val Gly Ile Ile 440 Ile Tyr Val Val Leu Leu Ile Phe Leu Lys Ala Glu Ile Ile Asn Lys 460 450 455 Leu Lys Phe Ile Met His Lys 470